

Biosynthesis of Phenanthroindolizidine Alkaloids *via* Derivatives of 2-Phenacetylpyrrolidine and Benzoylacetic Acid

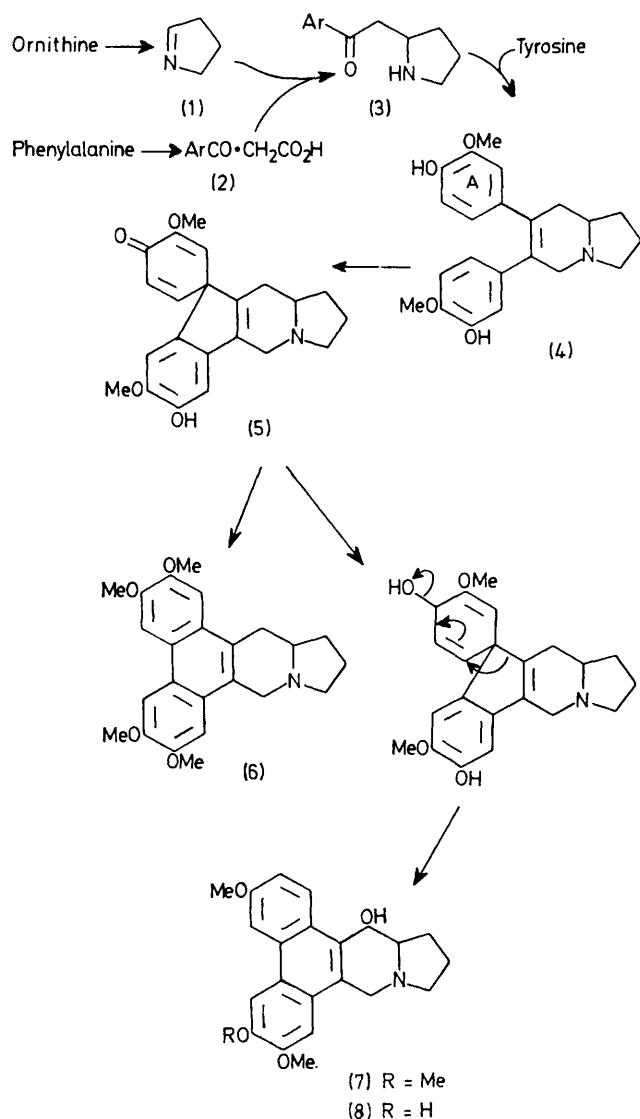
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Summary The derivatives of 2-phenacetylpyrrolidine, (**12**), (**13**), and (**14**), and of benzoylacetic acid, (**9**) and (**10**), are established as precursors of tylophorinine (**7**).

PREVIOUS results have shown that the phenanthroindolizidine alkaloid tylophorine (**6**) is derived from phenylalanine,¹ tyrosine,² and, probably, ornithine.^{1,3} A hypo-

thetical biosynthetic pathway (see Scheme) can be proposed (*cf.* refs. 1, 2, and 4) consonant with these observations and taking into account the structures of the

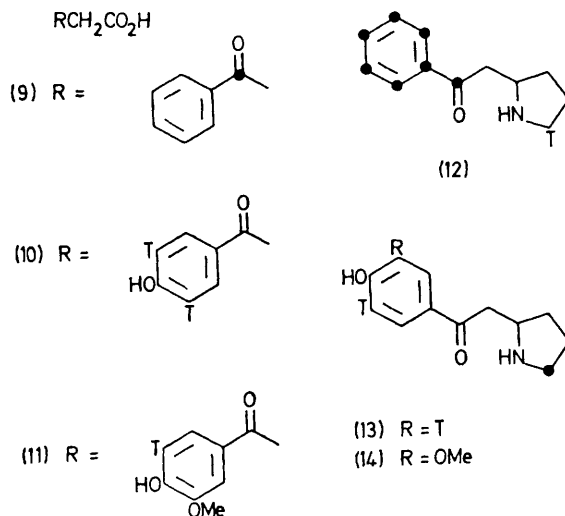


SCHEME

naturally occurring bases,⁵ represented by (4) and the six-membered-ring analogues of (3). Pivotal to such ideas are the 2-phenacylpyrrolidines (3) and we now report evidence that (12), (13), and (14) are intact precursors for tylophorinine (7) in *Tylophora asthmatica* Wight et Arn, thus establishing a biosynthetic pathway *via* these bases and giving substance to the previously held views. Further definition of tylophorinine biosynthesis was obtained by showing that (9) and (10) [*cf.* (2)] are also incorporated into (7).

2-Phenacylpyrrolidines (3) are conveniently prepared by condensation of Δ^1 -pyrrolidine (1) with the appropriately substituted benzoyl acetic acid.⁶ In this way samples of (12), (13), and (14) were synthesized with labels as shown

(^{14}C : ●). When they were administered to *T. asthmatica*, each of them was found to be a satisfactory precursor for tylophorinine (7) (Table). The amines (12) and (14) were incorporated without change in isotope ratio and are therefore utilized intact for biosynthesis. The incorporation of base (13) into tylophorinine (7) resulted in loss of essentially half the tritium. This was, however, expected since implication of (13) in the biosynthetic pathway logically involves entry of a hydroxy-group at one of the



tritiated sites. Thus (13) is also incorporated intact. These results strongly indicate that the biosynthesis of tylophorinine (7) proceeds by way of phenacylpyrrolidines (3) and involves in part the sequence represented as (12) → (13) → (14). Further, they imply that the biosynthesis of tylophorinine involves an intermediate (as 4) with a dioxygenated ring A, which is consistent with a pathway involving a dienone (5) which gives either tylophorine (6) by rearrangement, or tylophorinine (7) after reduction followed by rearrangement (*cf.* ref. 4).

TABLE. Incorporation of precursors into tylophorinine (7)^a

Precursor	Administered $^3\text{H}/^{14}\text{C}$	Isolated $^3\text{H}/^{14}\text{C}$	Isolated % Incorporation
(9) ^b	—	—	1.5×10^{-3}
(10) ^b	—	—	5.8×10^{-3}
(11) ^b	—	—	Inactive alkaloid
(12) ^c	12.6	13.0	1.7×10^{-1}
(13) ^b	6.1	2.6	3.8×10^{-3}
(14) ^b	2.7	3.3	3.5×10^{-2}

^a Isolated as its acetate. ^b Absorption into excised stems. ^c Wick-feed to whole plants.

Further support for, and definition of, the early part of the pathway shown in the Scheme comes from the finding that the keto-acids (9) and (10), but not (11) (labels as shown; ^{14}C : ●) were satisfactory precursors for tylophorinine (Table). Although the sites of labelling in the derived alkaloid were not established the observation that there was a marked distinction in the incorporation of label from (9) and (10) compared to (11) and that the levels of

incorporation of (9) and (10) were similar to those of (13) and (14) argues strongly that these keto-acids were incorporated intact. The incorporation of (10) indicates that hydroxylation of the aromatic nucleus can occur before formation of (3) as an alternative to hydroxylation of (12). However, the failure of (11) to act as a precursor suggests that only a single oxygen function may be introduced at the keto-acid level of biosynthesis.

A puzzling feature of our results was the isolation of inactive samples of tylophorine (6) and tylophorinidine (8) in all the experiments. In preliminary studies with tritiated samples of the precursors samples of (6), (7), and (8) were isolated with similar levels of radioactivity so we

presume that at the time of our later experiments neither (6) nor (8) were being biosynthesized.

In conclusion, our results establish the importance of the keto-acids (2) and keto-amines (3) in the biosynthesis of phenanthroindolizidine alkaloids as exemplified by tylophorinine (7), and point the way to further experiments notably with compounds such as (4). Further, it can be expected that 2-phenacylpyrrolidines (3) will be found in plants.

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